

Development of a New Green HPLC Method for Gatifloxacin Determination in Bulk Drug and Ophthalmic Formulations

Ola Mahmod Younes*¹

*¹ Doctor, applied analytical chemistry, Analytical and food chemistry department, Faculty of pharmacy Damascus university. Ola31.Younes@damascusuniversity.edu.sy

Abstract:

Background: Control of the active pharmaceutical ingredient plays a crucial role in ensuring the required efficacy. Therefore, a new accurate, reproducible, and green RP-HPLC method was developed for determining Gatifloxacin in bulk and eye drops.

Methods: The chromatographic separation was carried out using (isopropanol: phosphate buffer, pH=3±0.1) (35:65) v/v as mobile phase, the column used was C8 5μ (4.6*150mm) Inert Sustain GL Sciences column at temperature [40] °C, the flow rate was 0.8 mL/min, the UV detector was set at 280 nm and the injection volume was 10 μL. The greenness of the method was evaluated using Eco-scale tool.

Results: ICH guidelines were applied to prove the validity of the developed method. The linearity was achieved in the range 10-120 mg/l for Gatifloxacin with an excellent correlation coefficient R²=1. The LOD and LOQ were found to be 0.026 and 0.08 mg/l respectively. The precision was studied as intra- and inter-day with relative standard deviations not more than 2%, and the accuracy by mean recoveries ranged from 99.36 to 102.25%. The developed method was successfully applied for to determine Gatifloxacin in marketed eye drops. Eventually, the Eco-scale tool revealed a high degree of greenness by using relatively more eco-friendly solvents.

Conclusion: A new, sensitive, green, and accurate reversed-phase liquid chromatographic method is developed and validated to determine gatifloxacin in bulk and ophthalmic formulations using a UV detector under isocratic conditions.

Keywords: RP- HPLC, Gatifloxacin, Eye Drops, Method Validation, Greenness Assessment.



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تطوير طريقة كروماتوغرافية سائلة عالية الأداء جديدة خضراء لتحديد غاتيفلوكساسين كمادة أولية وفي الأشكال العينية

علا محمود يونس^{1*}

^{1*} مدرس- اختصاص الكيمياء التحليلية التطبيقية- قسم الكيمياء التحليلية والغذائية - كلية الصيدلة- جامعة دمشق.

Ola31.Younes@damascusuniversity.edu.sy

الملخص:

خلفية البحث وهدفه: تلعب مراقبة المواد الفعالة صيدلانياً دوراً هاماً في ضمان الفعالية المنشودة، لذلك تم تطوير طريقة جديدة، ودقيقة ومضبوطة وخضراء لتحديد غاتيفلوكساسين كمادة أولية وفي الأشكال العينية.

مواد البحث وطرائقه: تم اجراء الفصل الكروماتوغرافي باستعمال إيزوبروبانول: وقاء فوسفاتي (35: pH=3±0.1) (65/حجم/ حجم، كطور متحرك، عمود C8 بأبعاد 5 InertSustain GL Sciences (4.6*150mm) μ في درجة الحرارة [C40] °، كان معدل التدفق 0.8 مل / دقيقة، عند طول موجة 280 نانومتر، وحجم الحقنة 10 ميكرو لتر، تم تقييم خضرة الطريقة المطورة باستخدام أداة. Eco-Scale

النتائج: تم تطبيق إرشادات المؤتمر الدولي للتنسيق والمواءمة لإثبات مصدوقية الطريقة المطورة، كانت الخطية ضمن المجال 10-120 ملغ/ل من أجل غاتيفلوكساسين ومعامل ارتباط ممتاز قدره 1، وكان حدا الكشف والمقايضة الكمية 0.026، 0.08 ملغ/ل على الترتيب، تمت دراسة الدقة خلال اليوم الواحد وبين أيام مختلفة فكانت الانحرافات المعيارية النسبية لا تزيد عن 2٪، وتراوح المصبوطية بمتوسط استعادة من 99.36% حتى 102.25%. تم تطبيق الطريقة المطورة بنجاح لتحديد المركب في القطرات العينية التجارية. نهايةً، أظهرت نتيجة أداة Eco-Scale درجة عالية من الخضرة نظراً لاستخدام محلات أكثر صداقة للبيئة نسبياً.

الاستنتاج: تم تطوير طريقة كروماتوغرافية سائلة بالطور العكوس جديدة، حساسة، خضراء ودقيقة لتحديد غاتيفلوكساسين كمادة أولية وفي القطرات العينية، باستخدام التحري بالأشعة فوق البنفسجية تحت شروط النظام المتساير.

الكلمات المفتاحية: الكروماتوغرافيا السائلة عالية الأداء بالطور العكوس، غاتيفلوكساسين، القطرات العينية، مصدوقية الطريقة، تقييم خضرة الطريقة.

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1. Introduction:

Gatifloxacin (GTX) ((RS) 1-cyclopropyl-6-fluoro-1,4-dihydro-8-methoxy-7-(3-methylpiperazin-1-yl)-4-oxo-3-quinolinecarboxylic acid) (Figure1) [1] has broad antimicrobial activity and is effective for the treatment of a wide variety of infectious diseases [1]

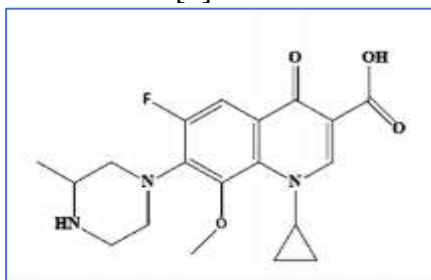


Figure 1: structure of Gatifloxacin[1]

Assay method for GTX has been cancelled in USP, however there is an official HPLC method in Indian Pharmacopoeia for Gatifloxacin determination as raw material and in ophthalmic solutions, infusions, and tablets [2].

A review of the literature on this drug had revealed chromatographic methods. Martin N Saad and his colleagues developed HPLC method for the simultaneous determination of Gatifloxacin and dexametasone in ophthalmic formulations using different column technologies including monolithic columns which showed the best results due to their hydrodynamic properties[3], another method was developed depending on ion pairing HPLC to determine GTX as bulk and in formulations[4], some HPLC methods have been conducted including reversed-phase HPLC method for drug determination in tablet dosage forms[5], stability-indicating HPLC method as bulk and in pharmaceutical preparations [6], HPLC method for the determination of Gatifloxacin stability in human plasma [7], on other

hand ,spectrophotometric assays directly or after derivation[8,9] and spectrofluorimetric methods based on charge transfer complex formation [10] or metal ion interaction [1] have been developed.

Most HPLC methods even Pharmacopoeial ones use acetonitrile or consume large quantities of methanol in the mobile phase. Unfortunately, such solvents are considered harmful to both analysts and environment. Moreover, some methods as mentioned above, use special technology of column. Because of HPLC technique is considered a worldwide approach for drug determination [11,12], this work aims at developing a new reversed-phase HPLC for GTX determination as bulk and in eye drops in a relatively short retention time by using green mobile phase, and classical C8 column, which is available in all analytical laboratories. The developed method was validated according to ICH guidelines. Among various greenness assessment tools that depend basically on the twelve principles of green analytical chemistry (waste prevention, atom economy maximizing, less hazardous chemical synthesis, safer chemical and products designing, use of renewable sources, avoid chemical derivation, use catalysts, design for degradation, real time pollution prevention, accidents prevention) [13], Eco-Scale calculator was employed.

2. Materials and methods

2.1. Instrumentation

Chromatographic separation was carried out using a Shimadzu HPLC system LC 2030C 3D plus (Shimadzu Corporation, Japan), equipped with a PDA detector, an autosampler, column oven (Shimadzu Corporation, Japan). C8 column 5 μ m (4.6*250mm) InertSustain GL Sciences Inc.

Japan. Ultrasonic bath -BANDELIN SONOREX, Balance A and D Company Limited, Japan. pH meter WTW-inolab pH7310p, PTFE syring filter 0.45 μ m.

2.2. Materials & standards

Potassium dihydrogen phosphate (KH_2PO_4) (99% purity), ortho phosphoric acid (85% purity) were purchased from HIMEDIA-INDIA. Isopropanol, HPLC grade from Scharlou-SPAIN. Standard Gatifloxacin sesquihydrate (99.9% purity) was kindly provided by Miamed Pharmaceutical Industries (Evergreen chemical factory Co; Ltd, China). Eye drops (containing Gatifloxacin 0.3%) were purchased from Syrian market.

2.3. Chromatographic conditions

InterSustain C8 Column 5 μ m (4.6 x250mm) (GL Sciences, Japan), temperature 40°C, and mobile phase consisted of (isopropanol: phosphate buffer, pH=3 \pm 0.1) (35 :65) v/v. Phosphate buffer consisted of 0.025M of KH_2PO_4 , the pH was adjusted by ortho phosphoric acid. The mobile phase was degassed by sonication for 15 min and filtered through 0.45 μ m Millipore membrane filter before use. (Figure. 2) shows the chromatogram of GTX under optimal conditions (flow rate 0.8 mL/min, and UV detection at 280 nm, injection volume 10 μ l).

2.4. Standard Solution

Stock solution of GTX was prepared at concentration of 500 mg/L, using mobile phase as a solvent and stored in refrigerator at 4°C temperature. Five working solutions were prepared by diluting the stock solution to obtain concentrations ranging from 10 to 120 mg/L..

2.5. Method validation

The developed method was validated according to the International Conference on Harmonization (ICH) guidelines, encompassing the following parameters: linearity, limit of detection (LOD), limit of quantification (LOQ), precision, accuracy, selectivity and robustness[14].

2.5.1. Linearity, limit of detection (LOD) and limit of quantification (LOQ)

It was evaluated by plotting calibration curve using five concentrations: 10-30-60-90-120 mg/L of GTX against area under the curve, each concentration was injected in triplicate. Slope, intercept and correlation coefficient R^2 of the calibration curve were calculated. LOD, LOQ were calculated based on LOD (3.3 SD/S) and LOQ (10 SD /S), where SD the standard deviation of the response and S the slope of the calibration curve.

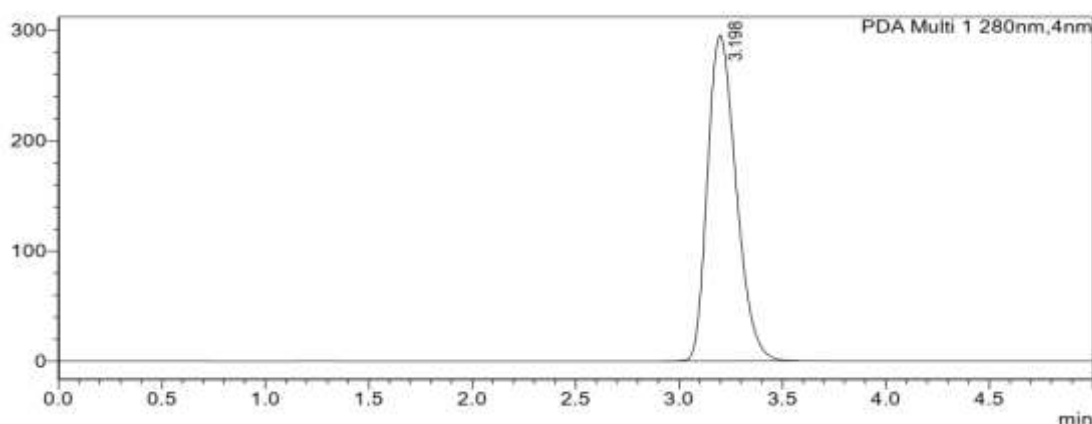


Figure 2: standard chromatogram of GTX by developed method

2.5.2. Precision

Two levels of precision were evaluated by applying the developed method on three concentrations on the same day (intra-day) and in three different days (inter-day), then relative standard deviation (RSD%) was calculated.

2.5.3. Accuracy

The accuracy of the developed method was evaluated using recovery and mean recovery at three different concentrations of GTX.

2.5.4. Robustness

Robustness was achieved by subjecting the method to a minor change in the conditions. Temperature was chosen as a parameter, then relative standard deviation (RSD %) was calculated.

2.5.5. Selectivity

The selectivity of the developed method was tested by applying the optimal chromatographic conditions to a mixture of common eye drop excipients (ethylenediaminetetraacetic acid EDTA, sodium chloride, benzalkonium chloride).

2.5.6. Determination of Gatifloxacin in marketed eye drop (0.3% GTX)

Equivalent to 0.6 g (2ml) of GTX was diluted to 100mL with mobile phase as a solvent, and after filtration, 10 μ l was

injected into the chromatographic system. Eventually, the practical content was calculated using a calibration curve equation.

3. Results and discussion

The first objective of this work is to replace acetonitrile with the minimum amount of safer solvent at the same time, using buffer solution as a maximum part of the mobile phase.

Since literature lacked examples of isopropanol as an organic modifier in mobile phases for Gatifloxacin analysis, this study investigated the optimal use of isopropanol alongside buffer to achieve accurate and reproducible Gatifloxacin determination.

3.1. Mobile phase composition

The optimal mobile phase composition, determined experimentally, consisted of 65% phosphate buffer and 35% isopropanol (Table 1). This composition effectively balanced rapid analyte elution with acceptable peak shapes.

The buffer pH was carefully adjusted to 3 to achieve symmetrical peak shapes and minimize retention times, thereby improving the overall chromatographic performance.

3.2. System suitability

System suitability parameters were calculated to ensure that, the applied

chromatographic system is acceptable under optimal conditions

Table 1: selection of mobile phase composition

Mobile phase compositi on	Parameters		t_R (min)	T	NTP
	Isopropanol%	Buffer %			
	10	90	12.600	1.50	3600
	20	80	3.870	-	2100
	35	65	3.195	1.30	2500
	40	60	2.580	overlapped	859

Table 2: System suitability

Retention time (t_R)	3.195
Tailing factor (T)	1.30
Capacity factor (K)*	2.04
Number of theoretical plates (NTP)	2500

* Dead time(t_m) = 1.05

3.3 method validation

3.3.1. Linearity, LOD, LOQ

The calibration curve demonstrated an excellent linear relationship within the range of 10–120 mg/L. Triplicate measurements were performed for each concentration, and the average peak areas were plotted against the corresponding concentrations. The correlation coefficient was determined to be 1, as illustrated in Figure 3. Using the previously defined equations, the limits of

detection (LOD) and quantification (LOQ) were calculated to be 0.026 mg/L and 0.08 mg/L, respectively

3.3.2. Accuracy and precision

To evaluate accuracy and precision, experiments were conducted at three different concentrations. The inter- and intra-day precision were assessed, yielding a relative standard deviation (RSD%) of less than 2. The detailed results are presented in Tables 3 and 4.

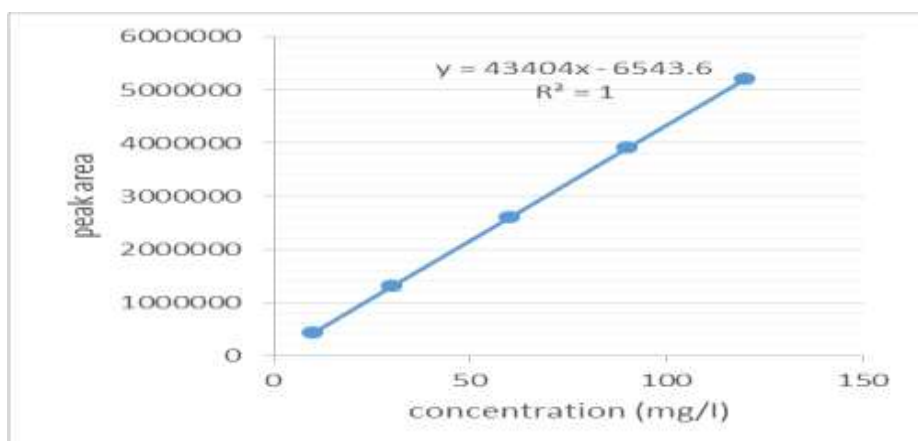


Figure 3: Linearity of the developed method

Table 3: Accuracy of the developed method

Concentration (mg/l)	Found (mg/l)*	Recovery %
20	20.45	102.25
60	59.62	99.36
120	120.66	100.55
		$Mean \pm SD = 100.72 \pm 1.45$

*Average of three separate determination

Table 4: Precision of the developed method

Concentration (mg/l)	intra – day*		inter – day*	
	Recovery \pm S.D*	Precision (RSD %)	Recovery% \pm S.D*	Precision (RSD %)
20	100.55 \pm 0.70	0.69	100.73 \pm 0.85	0.84
60	99.78 \pm 0.23	0.23	101.02 \pm 0.26	0.25
120	100.56 \pm 0.92	0.91	99.33 \pm 0.77	0.78

*Average of three separate determination

3.3.3. Robustness

To assess the robustness of the developed method, small variations in column

temperature were introduced. These changes had an insignificant effect on both peak area and recovery, as demonstrated in Table 5.

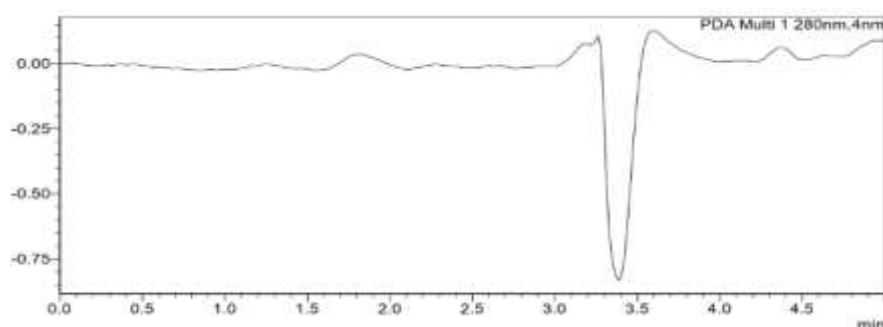
Table 5: Robustness of the developed method

(c) ⁰	38	40	42	Average% + SD	RSD %
recovery% of GTX	99.64	100.33	99.20	99.72% \pm 0.56	0.57

3.3.3. Selectivity

The method demonstrated excellent selectivity, as it was successfully applied in

the presence of excipients without any interference with the drug peak, as shown in the chromatogram in Figure 4.

**Figure 4: Selectivity of the developed method****3.4. Determination of Gatifloxacin in ophthalmic formulations**

The results of determination of GTX in marketed eye drops (0.3%) are summarized in table 6. The mean recovery was found to

be 100.74%, based on five replicate measurements. Figure 5 presents the chromatogram of GTX in the marketed ophthalmic solution.

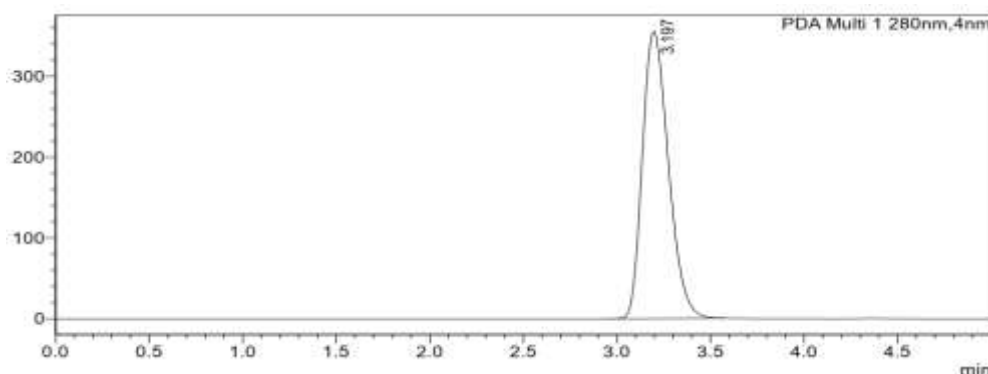


Figure 5: Chromatogram of GTX in marketed eye drops

Table 6: Determination of GTX in eye drops

Number of experiment	Recovery %	Average recovery % \pm SD
1	100.52	100.74 \pm 0.65
2	101.15	
3	99.98	
4	100.42	
5	101.64	

4.Greenness assessment using analytical Eco-Scale tool

The Analytical Eco-Scale is based on the principle that an ideal green analysis has a value of 100. For each parameter of the analytical procedure—such as reagent amounts, hazards, energy consumption, and waste generation—penalty points are assigned. The total penalty points for the procedure are used to calculate the Eco-Scale score using the following formula:

Analytical Eco-Scale = 100 - Total Penalty Points

The Eco-Scale score reflects the greenness of the method as follows:

- 75 indicates excellent green analysis.
- 50 indicates acceptable green analysis.
- 50< indicates inadequate green analysis [15].

As shown in Table 7, the developed method achieves an excellent level of greenness, reflecting its environmentally friendly attributes.

Table 7: Eco-Scale score of the developed method

Reagents	Penalty points (PPs)
Amount	1
Hazardous	2
$1 \times 2 \times 2 = 4$	
Instrumentation	
Energy	0
Occupational Hazard	3
Waste (amount)	3
Waste (management)	3
9	
$100 - (4+9) = 87$	

5. Conclusion:

Developing new analytical methods is a complex task, requiring the optimization of numerous parameters and conditions. In light of growing concerns about the future of our planet and its resources, the need for green analytical methods has become increasingly important. This study successfully developed and validated a sensitive, fast, precise, and environmentally friendly HPLC method for the determination of Gatifloxacin in bulk and ophthalmic solutions.

The main objective was to replace harmful solvents, such as acetonitrile, with greener alternatives in the mobile phase and to

incorporate a relatively high percentage of the aqueous component to minimize waste. The greenness of the method was assessed using the Analytical Eco-Scale tool, which demonstrated a high score. Based on these findings, the method is environmentally friendly and suitable for application in quality control laboratories.

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Statements and Declarations

Competing Interests :

- The corresponding author states that there is no conflict of interest.

