

Comparative evaluation of the antibacterial efficacy of sodium hypochlorite gel and solution at different concentrations against *Enterococcus faecalis* within the root canals of primary molars: an in vitro study

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Abstract:

Objectives:

The study aimed to evaluate the antibacterial efficacy of sodium hypochlorite (NaOCl) in its solution and gel forms, at different concentrations, as an irrigant against *Enterococcus faecalis* in primary molars.

Materials and methods:

The sample consisted of 50 upper and lower primary molars, divided into five equal groups: Group 1: 2.25% NaOCl solution. Group 2: 5.25% NaOCl solution. Group 3: 2.25% NaOCl gel. Group 4: 4% NaOCl gel. Group 5 (negative control): 0.9% saline. After irrigation, the teeth were incubated for 24 hours. A sample was then cultured on blood agar plates. The plates were then incubated for 72 hours, and the bacteria were counted.

Results:

NaOCl at all concentrations, both liquid and gel forms, demonstrated antibacterial activity against *Enterococcus faecalis*, with 2.25% NaOCl gel outperforming 4% ($p < 0.001$).

Conclusions

NaOCl gel at concentrations of 2.25%–4% can be considered an effective alternative against *Enterococcus faecalis* in the endodontic treatment of primary molars.

Key Words: Naocl Gel, Irrigants, Primary Molars, *Enterococcus Faecalis*.

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التقييم المقارن للفعالية المضادة للجراثيم لهلام ومحلول هيبوكلووريت الصوديوم بتركيزات مختلفة ضد المكورات المعوية البرازية داخل قنوات جذور الأرحاء المؤقتة: دراسة مخبرية

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الملخص:

الأهداف:

تهدف هذه الدراسة إلى مقارنة الفعالية المضادة للجراثيم لمحلول هيبوكلووريت الصوديوم 2.25% و 5.25% وهلام هيبوكلووريت الصوديوم 2.25% و 4% ضد المكورات المعوية البرازية في قنوات جذور الأرحاء المؤقتة.

المواد والطرائق:

تألقت العينة من 50 رحي مؤقتة مقسمة إلى خمس مجموعات متساوية: المجموعة 1: محلول 2.25% هيبوكلووريت الصوديوم. المجموعة 2: محلول 5.25% هيبوكلووريت الصوديوم. المجموعة 3: جل 2.25% هيبوكلووريت الصوديوم. المجموعة 4: جل 4% هيبوكلووريت الصوديوم. المجموعة 5 (مجموعة شاهدة سلبية): محلول ملحي 0.9%. حُضنت الأسنان لمدة 24 ساعة، ثم زُرعت العينة على أطباق الأغار الدموي وحُضنت لمدة 72 ساعة، ثم عُدت المكورات العقدية البرازية.

النتائج:

أظهر هيبوكلووريت الصوديوم بجميع تراكيزه، سواءً المحلول أو الهلامي، نشاطاً مضاداً للجراثيم ضد المكورات العقدية البرازية، حيث تفوق هلام هيبوكلووريت الصوديوم بتركيز 2.25% على تركيز 4% ($p < 0.001$).

الاستنتاجات:

يُمكن اعتبار هلام هيبوكلووريت الصوديوم بتركيزات تتراوح بين 2.25% و 4% بديلاً فعالاً ضد المكورات العقدية البرازية في المعالجة اللبية للأرحاء المؤقتة.

الكلمات المفتاحية: هلام هيبوكلووريت الصوديوم - سوائل الإرواء - الأرحاء المؤقتة - المكورات المعوية البرازية.

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حقوق النشر: جامعة دمشق -

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Introduction:

Dental caries is the most common chronic disease among pediatric patients [1] and is a common and increasing health problem, especially in developing countries [2]. The cause may be poor socioeconomic conditions and lifestyle changes. If caries is not treated appropriately, the most common consequences are pain, infection, premature tooth loss, loss of dental arch length, decreased chewing efficiency, impacted permanent teeth, and loss of self-confidence [3]. Endodontic treatment is indicated when caries affects the pulp and if infection has reached the root pulp [4]. The primary goal of root canal treatment is to remove bacteria and their products and infected tissue from the root canal system through disinfection and chemical and mechanical debridement [5].

Bacteria play a major role in the initiation and progression of endodontic and periapical diseases [6]. Mechanical debridement of the root canals alone cannot eliminate bacteria from the primary root canals due to their complex anatomy [7]. Therefore, reliance on irrigants becomes even more important for removing bacteria in the primary canals of teeth with their thin root dentin walls, complex internal anatomy, and irregular root canal system [8].

Sodium hypochlorite (NaOCl) solution is the most widely used irrigant. Despite its unpleasant taste and pungent odor, it has a broad spectrum of bactericidal activity [9]. NaOCl solution is used in concentrations ranging from 0.5% to 6% [10]. It is good at dissolving pulpal debris, collagen fibers, and dead tissue. It also provides complete debridement of the root canals, lubrication is easy to maintain, and has a long shelf life [11].

One of its major drawbacks is its high toxicity to living tissue, especially if the irrigant escapes from the apex, which can cause significant damage to periapical tissues or trigger an allergic reaction [12]. Applying NaOCl in gel form offers better control over application and reduces the risk of over-apexing, especially in children with young and primary teeth. The gel also has a more pleasant odor and taste than the liquid [13, 14]. This study aims to compare the antibacterial efficacy of 2.25% and 5.25% NaOCl Solution and 2.25% and 4% NaOCl gel against *Enterococcus faecalis* in the root canals of primary molars. The null hypothesis is that no statistically significant difference would be noted

between NaOCl solution and gel at different concentrations in the antibacterial efficacy against *Enterococcus faecalis*.

Materials And Methods:

Study design and ethics:

It was an in vitro study conducted at the Pediatric Dentistry Department of the Faculty of Dentistry at Damascus University in September 2024. The Local Ethics Committee at Damascus University approved the study under N/2024, and the research adhered to the Checklist for Reporting In-Vitro Studies (CRIS) guidelines [15]. Written informed consent for tooth donation was obtained from the legal guardians of the patients, and the extraction of human primary molars was carried out for pathological or orthodontic purposes.

Sample size calculation and tooth selection:

The criteria for determining the sample size included an effect size of 0.51 (effect size $f=0.51$), a two-tailed significance level of 5% ($\alpha=0.05$), a 95% confidence interval, a statistical power of 80% ($1-\beta$ err prob = 0.80), and five experimental groups. Fifty specimens were gathered for the sample size, and the effect size calculation was derived from an initial study [16]. Teeth exhibiting root resorption greater than one-third of the root length, those that have had prior endodontic treatment, roots with calcified canals, and teeth presenting fractures or fissures were excluded. All teeth were subjected to disinfection by being immersed in a 5% NaOCl solution (Carmel®; AkkaBrothers Co., Carmel Detergent, Damascus, Syria) for one hour. Following the disinfection process, the teeth were thoroughly rinsed with tap water and preserved in normal saline (SODIUM CHLORIDE 0.9% MIAMED, Miamed Pharmaceutical Industry, Damascus, Syria) until utilized for the study [17]. The fifty human primary molars were randomly allocated into five groups ($n = 10$):

- Group 1: irrigation with 2.25% NaOCl solution (Hypo Chlorite, Noura, Damascus, Syria).
- Group 2: irrigation with 5.25% NaOCl solution.
- Group 3: irrigation with 2.25% NaOCl gel (BLEACH gel, WC NET, Milano, Italy).

- Group 4: irrigation with 4% NaOCl gel (LET'S CLEAN Concentrated Chlorine, DTIC®, Damascus, Syria).
- Group 5 (negative control): irrigation with 0.9% saline.

Randomization and blinding

The sample was randomly assigned using a lottery method, with numbers from 1 to 50 written on paper cards. The study was double-blinded, where the outcome assessor and the statistician were unaware of the groups.

Procedure

The access cavity was performed. The working length was determined using a 15 K-file size (K File,

FANTA DENTA®, Shanghai, China) until the file penetrated and retreated 1 mm. A 20 K-file size 4% taper (SC Niti File, SOCO PRECISION INSTRUMENT CO. LTD., Guangdong, China) was used, followed by a 1 mL irrigation with saline (SODIUM CHLORIDE 0.9% MIAMED, Miamed Pharmaceutical Industry, Damascus, Syria). Next, A 25 K-file size 4% taper was used, followed by a 1 mL irrigation with 5% NaOCl solution for 3 min, followed by a final irrigation with saline. Teeth were placed in acrylic cubes for ease of handling. The canals were dried, then moist-heat sterilized at 121°C for 20 min (Figure 1) [18].



Figure(1): Study sample within moist heat sterilization bags.

Enterococcus faecalis was cultured on blood agar plates every few days to ensure fresh bacteria were used (Figure 2). A suspension containing *Enterococcus faecalis* at a concentration of 1.5×10^8 CFU/mL, equivalent to 0.5 on the McFarland scale,

was prepared using a turbidimeter (TB1 Turbidimeter, A-MATRIX®, Lagos State, Nigeria). The sample was shaken on a shaker to ensure the solution was homogeneous. The glass tube was then placed in the turbidimeter [19].



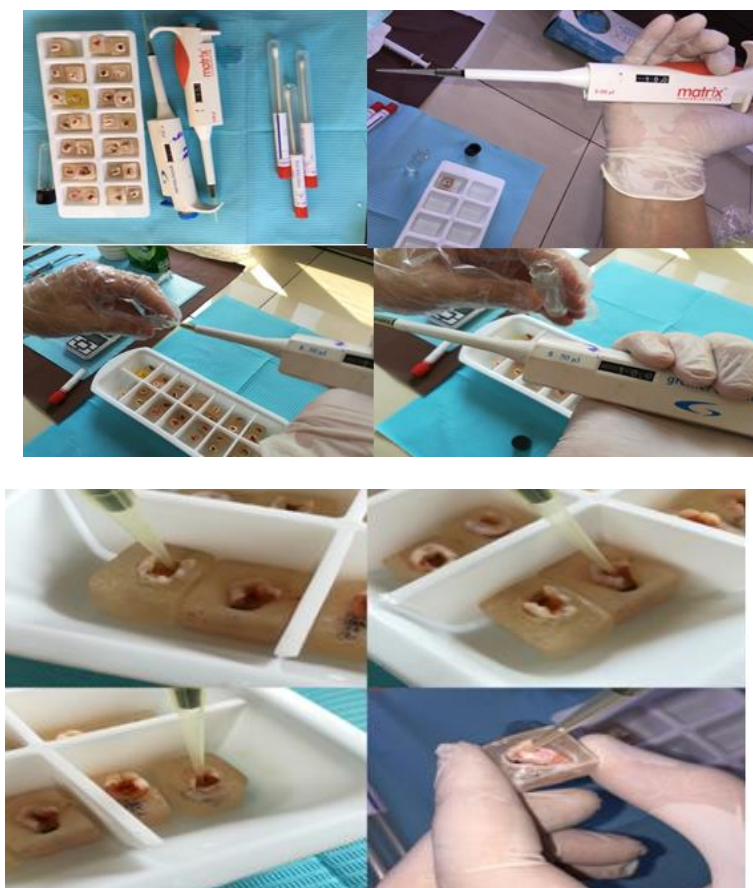
Figure(2): Cultivation of *Enterococcus faecalis* and use of fresh bacteria.

The experiment was repeated until a concentration of 0.5 McFarland was reached. Ten μ L of the bacterial suspension was taken using a micropipette (Micropipette 100, Matrix Healthcare Pvt. Ltd., New Delhi, India) and placed in the root canals (Figure 3). The samples were then incubated in an anaerobic incubator for 24 hours. Root canals were irrigated with 5 mL of each irrigant for 5 minutes using a 31-

gauge side-vented irrigating needle. They were then flushed with 5 ml of sterile saline. The root canals were dried along their entire length, filled with sterile saline, and circumferentially filed with an H-file for 15 seconds. Bacterial swabs were then collected using paper cones. The paper cone was inserted and left for 60 seconds before being transferred into an Eppendorf tube containing 2 mL

of sterile saline. The swabs were repeated three times to transfer the bacterial count, and the tube containing the paper cones was shaken for 1 min to ensure solution homogeneity. Ten μL of the solution was taken using a micropipette and cultured on pre-

prepared Petri plates, and placed in an incubator at 37°C . After 72 hours, the plates were removed from the incubator, and the bacterial colonies were counted. The units were converted to logarithmic numbers to facilitate statistical analysis.



Figure(3): Contamination of the root canals with Enterococcus faecalis.

Statistical analysis

IBM SPSS software version 26 (IBM Corp., Armonk, NY, USA) was utilized for statistical analysis. One-way ANOVA test was used to compare the mean differences in mean logarithmic reduction between study groups, and the Bonferroni test was used to study pairwise differences between groups. The logarithmic reduction of bacterial colony counts was calculated according to the following equation:

$$\text{Log reduction} = \log_{10}(\text{A}) - \log_{10}(\text{B})$$

A: The bacterial count before irrigation.

B: The bacterial count after irrigation.

Results:

Descriptive statistics of the decimal logarithm of bacterial count after irrigation is listed in Table 1.

Table 2 shows a statistically significant difference in the mean logarithmic reduction between the study groups ($p < 0.001$). Therefore, the null hypothesis was rejected. Table 3 shows a statistically significant difference between the 0.9% saline group and other study groups ($p < 0.001$) and between the 4% NaOCl gel group and 5.25% NaOCl solution and 2.25% NaOCl gel groups ($p < 0.001$) (Figure 4 and Figure 5).

Table(1): Descriptive statistics of the decimal logarithm of bacterial count after irrigation.

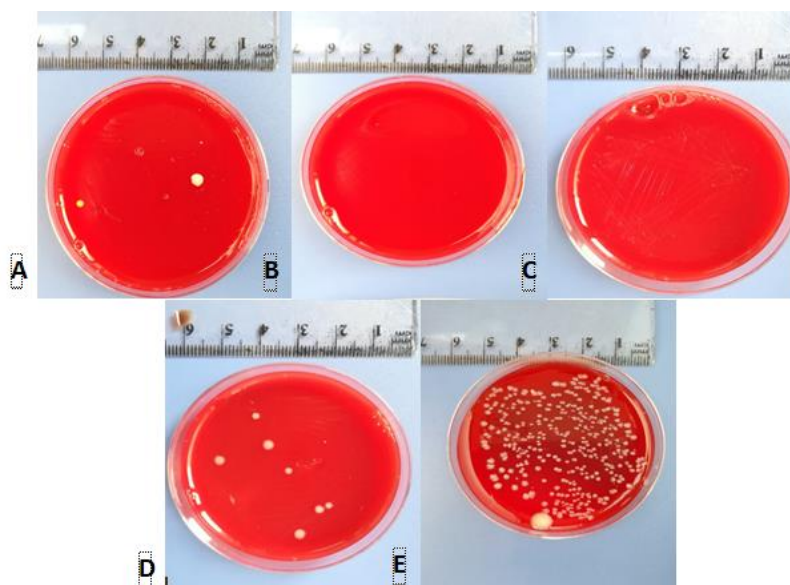
Groups	Mean	SD	Min	Max
2.25% NaOCl solution	1.04	1.15	0	1.66
5.25% NaOCl solution	0.30	0.48	0	0.95
2.25% NaOCl gel	0.30	0.48	0	0.90
4% NaOCl gel	1.85	1.93	0.48	2.43
0.9% saline	4.60	4.32	3.61	4.84

Table(2): One-way Anova test results for logarithmic reduction of bacterial counts.

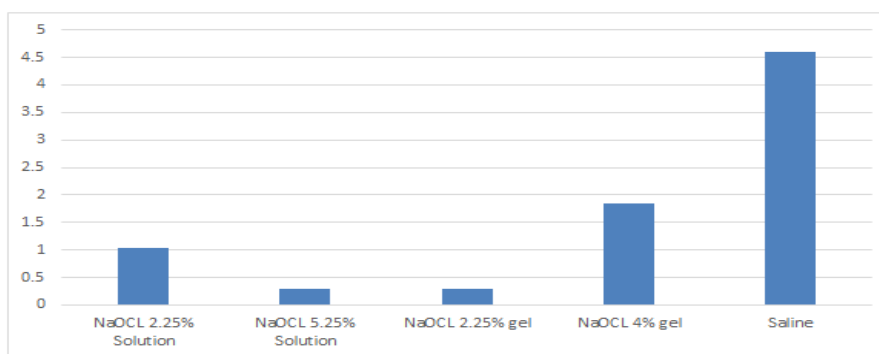
Groups	<i>p</i> -value
2.25% NaOCl solution	< 0.001*
5.25% NaOCl solution	
2.25% NaOCl gel	
4% NaOCl gel	
0.9% saline	

Table(3): Bonferroni test results for pairwise comparisons of logarithmic decimal reduction of bacterial counts.

Pairwise comparisons	<i>p</i> -value	
2.25% NaOCl solution	5.25% NaOCl solution	0.104
	2.25% NaOCl gel	0.158
	4% NaOCl gel	0.167
	0.9% saline	0.001*
5.25% NaOCl solution	2.25% NaOCl gel	0.830
	4% NaOCl gel	0.003*
	0.9% saline	< 0.001*
2.25% NaOCl gel	4% NaOCl gel	0.004*
	0.9% saline	< 0.001*
4% NaOCl gel	0.9% saline	0.043*



Figure(4): The bacterial count remaining on blood agar plates after irrigation. (A) 2.25% NaOCl solution. (B) 5.25% NaOCl solution. (C) 2.25% NaOCl gel. (D) 4% NaOCl gel. (E) 0.9% saline.



Figure(5): The decimal logarithm of bacterial count after irrigation

Discussion:

Necrotic primary teeth contain numerous microorganisms, particularly anaerobic bacteria, which reside deep within the dentin canals and cementum around the apical region. These microorganisms persist even after mechanical preparation of the root canals. Mechanical preparation alone cannot effectively eliminate bacteria in the canals of primary teeth due to their anatomical complexity [20]. Therefore, reliance on irrigation has become increasingly important, especially in primary teeth due to their thin walls and irregular root canal anatomy [8]. NaOCl solution is considered the best irrigating solution for root canals. It possesses antibacterial activity by releasing hypochlorous acid (HClO), which exerts an oxidative action on the sulfhydryl groups of bacterial enzymes, disrupting bacterial cell metabolism [21]. The current study compares the antibacterial activity of 5.25% and 2.25% NaOCl solution and 4% and 2.25% NaOCl gel against *Enterococcus faecalis* in vitro on maxillary and mandibular primary molars.

Enterococcus faecalis was chosen because it is one of the most resistant intracanal bacteria and the most common bacteria isolated from periapical infections. *Enterococcus faecalis* is the most dominant bacterium in failed root canal treatments, accounting for 50-70% of these cases. It is due to its high survival capacity, resistance to antiseptics used in root canals, and ability to survive in low-nutrient environments. The concentration of the *Enterococcus faecalis* suspension was selected at 1.5×10^8 CFU/mL, according to the study of Öter et al. [19]. The suspension was introduced using a micropipette with atomized tips to ensure the bacterial sample was not contaminated with other bacteria.

The study sample consisted of 50 maxillary and mandibular primary molars. Considering that the anatomy and resorption of the root canals can affect the bacterial load, the palatal roots of the maxillary second molars and the distal roots of the mandibular second molars were used to standardize the samples. Furthermore, these roots have a higher curvature than the maxillary and mesial canals [22]. The irrigation tip was 1 mm away from the full root length, according to a study of Pinker et al. [23]. The irrigation protocol was standardized in terms of time and volume of irrigant. Irrigation time ranged from 30 seconds to 10 min, while the volume of irrigant ranged from 0.05 mL to 15 mL. Irrigation was performed with 5 mL of the irrigant used in this study. Due to the different viscosities of the irrigant fluids and the pressure applied to the syringe, the irrigation speed was standardized across all groups at 1 mm per minute [24].

A vibrating device was used to shake the tubes for one min to ensure homogeneity of the suspension before transferring them to the blood agar plate. Ten μ L of the suspension were then withdrawn using a micropipette with a plastic tip for each sample to ensure no contamination. The suspension was spread on the plate using a culture tube so that it was distributed over the entire surface of the plate.

NaOCl was used in this study because it is considered the gold standard irrigant. It can dissolve dead pulp tissue and collagen, and is readily available and inexpensive. The 5.25% NaOCl solution concentration was chosen for this study because it is the most common concentration used in endodontic treatments. The 2.25% NaOCl solution concentration was chosen to resemble the gel concentration used in this study. It is close to the 2.5% concentration, which eliminates toxins produced, and because of its ability to dissolve organic tissue [9-11]. Vaziri et

al. [25] also demonstrated that a 2.5% NaOCl solution was effective against *Enterococcus faecalis* upon direct contact after two min.

The results of this study are consistent with the study by Abu Hasna et al. [26], who evaluated the effectiveness of 2.5% NaOCl solution and 3% NaOCl gel against *Enterococcus faecalis*. The results demonstrated no statistically significant differences in the antibacterial effect of the solution and gel forms. They are also consistent with the study by Zand et al. [27], who indicated that both the 2.25% gel and the 2.25% and 5.25% solutions were able to eliminate *Enterococcus faecalis*, but the gel demonstrated a lower efficacy compared to the solution at both concentrations. The lower antibacterial efficacy of the gel compared to the solution is likely due to differences in the manufacturers of the gels used in their study. The results of this study were consistent with the study Sahebi et al. [6], as there was no statistically significant difference between the 2.25% NaOCl gel and liquid, and both were

effective in eliminating bacteria. All concentrations of NaOCl gel and solution showed antibacterial activity, with a statistically significant difference compared to the saline group, which did not show any antibacterial activity but had a rinsing effect that contributed to reducing the number of bacteria.

Conclusions:

Within the limitations of this study, we recommend the use of NaOCl gel at concentrations of 2.25% (WC NET) and 4% (LETS CLEAN) for 5 min in endodontic treatment of primary molars in children. It is considered a safe and effective alternative to solution against *Enterococcus faecalis*, with preference given to the 2.25% NaOCl WC NET gel.

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References:

1. Chen, X., Liu, X. and Zhong, J. (2017) 'Clinical and radiographic evaluation of pulpectomy in primary teeth: A 18-months clinical randomized controlled trial', *Head and Face Medicine*, 13(1). doi: 10.1186/s13005-017-0145-1.
2. Bahaa Aldin Alhaffar, M. H. et al. (2018) 'Seven years of war in Syria: The relation between oral health and PTSD among children', *Indian Journal of Oral Health and Research*, 4(1). doi: 10.4103/ijohr.ijohr_8_18.
3. Rasidi, M. Q. Z. B. M., Bhagya Lakshmi, T. and Prabu, D. (2020) 'Pulpectomy in maxillary first molars with distal caries', *International Journal of Research in Pharmaceutical Sciences*, 11(Special Issue 3). doi: 10.26452/ijrps.v11iSPL3.2962.
4. Makarem, A., Ravandeh, N. and Ebrahimi, M. (2014) 'Radiographic assessment and chair time of rotary instruments in the pulpectomy of primary second molar teeth: a randomized controlled clinical trial.', *Journal of Dental Research, Dental Clinics, Dental Prospects*, 8(2). doi: 10.5681/joddd.2014.015.
5. Imani, Zhila et al. (2018) 'Antibacterial effects of chitosan, formocresol and CMCP as pulpectomy medicament on enterococcus faecalis, staphylococcus aureus and streptococcus mutans', *Iranian Endodontic Journal*, 13(3). doi: 10.22037/iej.v13i3.20791.
6. Sahebi, S. et al. (2014) 'Comparison of the antibacterial effect of sodium hypochlorite and aloe vera solutions as root canal irrigants in human extracted teeth contaminated with enterococcus faecalis.', *Journal of dentistry (Shiraz, Iran)*, 15(1).
7. Al-Aloush, B. R., and Bshara, N. (2013). 'A Descriptive Study for the Root Canals in Primary Molars and their ramifications-in vitro study.', *Damascus University Journal for Health Sciences*, 29(1), 521-531.
8. Kumar, V. M. et al. (2014) 'The antimicrobial effectiveness of 25% propolis extract in root canal irrigation of primary teeth', *Journal of Indian Society of Pedodontics and Preventive Dentistry*, 32(2). doi: 10.4103/0970-4388.130786.
9. Moradi, F. and Haghgoo, R. (2018) 'Evaluation of antimicrobial efficacy of nanosilver solution, sodium hypochlorite and normal saline in root canal irrigation of primary teeth', *Contemporary Clinical Dentistry*, 9(6). doi: 10.4103/ccd.ccd_95_18.
10. Forghani, M. et al. (2017) 'Effect of a passive sonic irrigation system on elimination of Enterococcus faecalis from root canal systems of primary teeth, using different concentrations of sodium hypochlorite: An in vitro evaluation', *Journal of Dental Research, Dental Clinics, Dental Prospects*, 11(3). doi: 10.15171/joddd.2017.032.
11. Kapdan, A. et al. (2015) 'Which is the most effective disinfection method in primary root canals: Conventional or newly developed ones?', *Nigerian Journal of Clinical Practice*, 18(4). doi: 10.4103/1119-3077.154207.
12. Sundaram, D., Narayanan, R. K. and Vadakkepurayil, K. (2016) 'A comparative evaluation on antimicrobial effect of honey, neem leaf extract and sodium hypochlorite as intracanal irrigant: An ex-vivo study', *Journal of Clinical and Diagnostic Research*, 10(8). doi: 10.7860/JCDR/2016/19268.8311.
13. Al Nesser, S. F. and Bshara, N. G. (2019) 'Evaluation of the apical extrusion of sodium hypochlorite gel in immature permanent teeth: An in vitro study', *Dental and Medical Problems*, 56(2). doi: 10.17219/dmp/103911.
14. Karkoutly, M., and Bshara, N. (2022). 'Comparative evaluation of apical extrusion of sodium hypochlorite gel and solution in primary molars using two different instrumentation techniques: an in-vitro study.', *Journal of Stomatology*, 75(4), 238-244.
15. Krithikadatta, J., Gopikrishna, V., and Datta, M. (2014). 'CRIS Guidelines (Checklist for Reporting In-vitro Studies): A concept note on the need for standardized guidelines for improving quality and transparency in reporting: in-vitro: studies in experimental dental research.', *Journal of Conservative Dentistry*, 17(4), 301-304.
16. Berben, L., Sereika, S. M., and Engberg, S. (2012). 'Effect size estimation: methods and examples.', *International journal of nursing studies*, 49(8), 1039-1047.
17. Lee, B. S., et al. (2004). 'The role of organic tissue on the punch shear strength of human dentin.', *Journal of dentistry*, 32(2), 101-107.

18. Sandino-Lacayo, K. L., et al. (2024). 'Cyclic fatigue resistance of two pediatric rotary files manufactured with different heat treatments: an invitro study.', *J Clin Pediatr Dent*, 48(5), 102-9.
19. Öter, B. et al. (2018) 'Evaluation of Antibacterial Efficiency of Different Root Canal Disinfection Techniques in Primary Teeth', *Photomedicine and Laser Surgery*, 36(4). doi: 10.1089/pho.2017.4324.
20. Pinky, C., Shashibhushan, K. K. and Subbareddy, V. V. (2011) 'Endodontic treatment of necrosed primary teeth using two different combinations of antibacterial drugs: An in vivo study', *Journal of Indian Society of Pedodontics and Preventive Dentistry*, 29(2). doi: 10.4103/0970-4388.84684.
21. Tsai, C. F., Chung, J. J., Ding, S. J., and Chen, C. C. (2024). 'In vitro cytotoxicity and antibacterial activity of hypochlorous acid antimicrobial agent.', *Journal of Dental Sciences*, 19(1), 345-356.
22. Katge, F. and Wakpanjar, M. M. (2018) 'Root canal morphology of primary molars by clearing technique: An in vitro study', *Journal of Indian Society of Pedodontics and Preventive Dentistry*, 36(2). doi: 10.4103/JISPPD.JISPPD_237_16.
23. Pinker, M., Frank, W., Wrbas, K. T., and Tchorz, J. P. (2024). 'Influence of Root Canal Size and Curvature on Insertion Depth of Three Different Endodontic Irrigation Needles.', *Oral*, 4(4), 459-466.
24. Alkhourbotly, D., et al. (2022). 'Evaluation of the Antibacterial efficacy of QMix and AgNP solutions in Root canals of primary molars: an In-Vitro Study.', *Cureus*, 14(9).
25. Vaziri, S. et al. (2012) 'Comparison of the bactericidal efficacy of photodynamic therapy, 2.5% sodium hypochlorite, and 2% chlorhexidine against *Enterococcus faecalis* in root canals; an in vitro study', *Dental Research Journal*, 9(5). doi: 10.4103/1735-3327.104882.
26. Abu Hasna, A. et al. (2020) 'Effect of sodium hypochlorite solution and gel with/without passive ultrasonic irrigation on *Enterococcus faecalis*, *Escherichia coli* and their endotoxins', *F1000Research*, 9. doi: 10.12688/f1000research.24721.1.
27. Zand, V. et al. (2016) 'Antibacterial efficacy of different concentrations of sodium hypochlorite gel and solution on *Enterococcus faecalis* biofilm', *Iranian Endodontic Journal*, 11(4). doi: 10.22037/iej.2016.11.